

**COMPOSITIONS AND METHODS FOR DIAGNOSING,
MONITORING, STAGING, IMAGING AND TREATING STOMACH CANCER**

INTRODUCTION

This application claims the benefit of priority from
5 U.S. Provisional Application Serial No. 60/193,095, filed
March 30, 2000.

FIELD OF THE INVENTION

This invention relates, in part, to newly identified
polynucleotides and polypeptides encoded thereby, as well as
10 methods for producing and using these polynucleotides and
polypeptides. Antibodies which are immunospecific for these
polypeptides are also described. Expression of the newly
identified polynucleotides and levels of the polypeptides
encoded thereby are upregulated in or specific to stomach
15 cancer tissue. These new polynucleotides and polypeptides,
referred to herein as Stomach Cancer Specific Genes or SSGs
are believed to be useful in assays for detecting, diagnosing,
monitoring, staging, prognosticating, imaging and treating
cancers, particularly stomach cancer.

20 BACKGROUND OF THE INVENTION

Cancer of the stomach, also referred to as gastric
cancer, is difficult to diagnose in early stages and can be
in the stomach for a long time, growing to a large size before
symptoms arise. In the early stages of cancer of the stomach,
25 an individual may experience indigestion and stomach
discomfort, a bloated feeling after eating, mild nausea, loss
of appetite or heartburn. In more advanced stages of stomach
cancer, there may be blood in the stool, vomiting, weight loss
or more severe pain.

Patients diagnosed with early stage cancer generally have a much greater five-year survival rate as compared to the survival rate for patients diagnosed with distant metastasized cancers. Accordingly, new diagnostic methods which are more
5 sensitive and specific for detecting early cancer of the stomach are clearly needed.

Further, patients with gastrointestinal cancers such as stomach cancer are closely monitored following initial therapy and during adjuvant therapy to determine response to therapy
10 and to detect persistent or recurrent disease of metastasis. Thus, there is clearly a need for cancer markers which are more sensitive and specific in detecting recurrence of stomach cancer.

Another important step in managing stomach cancer is to
15 determine the stage of the patient's disease. Stage determination has potential prognostic value and provides criteria for designing optimal therapy. Generally, pathological staging of cancer is preferable over clinical staging because the former gives a more accurate prognosis.
20 However, clinical staging would be preferred were it at least as accurate as pathological staging because it does not depend on an invasive procedure to obtain tissue for pathological evaluation. Staging of stomach cancer would be improved by identifying new markers in cells, tissues, or bodily fluids
25 which could differentiate between different stages of invasion.

The present invention relates to newly identified polynucleotides and polypeptides encoded thereby which are referred to herein as Stomach Cancer Specific Genes or SSGs,
30 as well as antibodies which are immunospecific for the polypeptides. The present invention also relates to methods for use of these SSGs in detecting, diagnosing, monitoring, staging, prognosticating, imaging and treating stomach cancer. For purposes of the present invention, SSG refers, among other
35 things, to native protein expressed by the gene comprising a

polynucleotide sequence of SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 or 13. By SSG it is also meant herein polynucleotides which, due to degeneracy in genetic coding, comprise variations in nucleotide sequence as compared to SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 or 13, but which still encode the same protein. In the alternative, what is meant by SSG as used herein, means the native mRNA encoded by the gene comprising the polynucleotide sequence of SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 or 13, or a polynucleotide which is capable of hybridizing under stringent conditions to the antisense sequence of SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 or 13.

Other objects, features, advantages and aspects of the present invention will become apparent to those of skill in the art from the following description. It should be understood, however, that the following description and the specific examples, while indicating preferred embodiments of the invention are given by way of illustration only. Various changes and modifications within the spirit and scope of the disclosed invention will become readily apparent to those skilled in the art from reading the following description and from reading the other parts of the present disclosure.

SUMMARY OF THE INVENTION

Toward these ends, and others, it is an object of the present invention to provide isolated polynucleotide sequences comprising SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 or 13; fragments or portions of such sequences which contain at least 15 contiguous nucleobases of SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 or 13; nucleic acid sequences which, due to degeneracy in genetic coding, comprise variations in polynucleotide sequence as compared to SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 or 13, but which still encode the same protein; and nucleic acid sequences which are capable of hybridizing under stringent conditions to the antisense

sequence of SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12
or 13. The present invention further relates to isolated
polypeptides encoded by the above-described polynucleotides.
These polynucleotides and/or the polypeptides encoded thereby
5 are referred to generally herein as Stomach Cancer Specific
Genes or SSGs.

It is another object of the present invention to provide
vectors containing the SSG polynucleotides and host cells for
expression and recovery of the SSG polypeptides encoded
10 thereby.

It is another object of the present invention to provide
antibodies which are immunospecific for SSG polypeptides.

It is another object of the present invention to provide
tools for detection of SSGs. Such tools include, but are not
15 limited to, antisense oligonucleotides which specifically
hybridize with the SSGs and antibodies which are
immunospecific for the SSGs.

It is another object of the present invention to provide
a method for diagnosing the presence of stomach cancer by
20 analyzing for changes in levels of SSG in cells, tissues or
bodily fluids compared with levels of SSG in preferably the
same cells, tissues, or bodily fluid type of a normal human
control, wherein a change in levels of SSG in the patient
versus the normal human control is associated with stomach
25 cancer.

Further provided is a method of diagnosing metastatic
cancer in a patient having stomach cancer which is not known
to have metastasized by identifying a human patient suspected
of having stomach cancer that has metastasized; analyzing a
30 sample of cells, tissues, or bodily fluid from such patient
for SSG; comparing the SSG levels in such cells, tissues, or
bodily fluid with levels of SSG in preferably the same cells,
tissues, or bodily fluid type of a normal human control,
wherein an increase in SSG levels in the patient versus the

normal human control is associated with stomach cancer which has metastasized.

Also provided by the invention is a method of staging stomach cancer in a human which has such cancer by identifying
5 a human patient having such cancer; analyzing a sample of cells, tissues, or bodily fluid from such patient for SSG; comparing SSG levels in such cells, tissues, or bodily fluid with levels of SSG in preferably the same cells, tissues, or bodily fluid type of a normal human control sample, wherein
10 an increase in SSG levels in the patient versus the normal human control is associated with a cancer which is progressing and a decrease in the levels of SSG is associated with a cancer which is regressing or in remission.

Further provided is a method of monitoring stomach
15 cancer in a human having such cancer for the onset of metastasis. The method comprises identifying a human patient having such cancer that is not known to have metastasized; periodically analyzing a sample of cells, tissues, or bodily fluid from such patient for SSG; comparing the SSG levels in
20 such cells, tissue, or bodily fluid with levels of SSG in preferably the same cells, tissues, or bodily fluid type of a normal human control sample, wherein an increase in SSG levels in the patient versus the normal human control is associated with a cancer which has metastasized.

Further provided is a method of monitoring the change
25 in stage of stomach cancer in a human having such cancer by looking at levels of SSG in a human having such cancer. The method comprises identifying a human patient having such cancer; periodically analyzing a sample of cells, tissues, or
30 bodily fluid from such patient for SSG; comparing the SSG levels in such cells, tissue, or bodily fluid with levels of SSG in preferably the same cells, tissues, or bodily fluid type of a normal human control sample, wherein an increase in SSG levels in the patient versus the normal human control is
35 associated with a cancer which is progressing and a decrease

in the levels of SSG is associated with a cancer which is regressing or in remission.

Further provided are new therapeutic agents and methods of identifying therapeutic agents targeted to SSGs for use in
5 imaging and treating disease relating to SSGs such as stomach cancer. For example, in one embodiment, therapeutic agents such as antibodies targeted against SSG or fragments of such antibodies can be used to detect or image localization of SSG in a patient for the purpose of detecting or diagnosing a
10 disease or condition. Such antibodies can be polyclonal, monoclonal, or omniclonal or prepared by molecular biology techniques. The term "antibody", as used herein and throughout the instant specification, is also meant to include aptamers and single-stranded oligonucleotides such as those
15 derived from an *in vitro* evolution protocol referred to as SELEX and well known to those skilled in the art. Antibodies can be labeled with a variety of detectable labels including, but not limited to, radioisotopes and paramagnetic metals. Therapeutic agents such as small molecules or antibodies or
20 fragments thereof which decrease the concentration and/or activity of SSG can also be used in the treatment of diseases characterized by expression of SSG. In these applications, the antibody can be used without or with derivatization to a cytotoxic agent such as a radioisotope, enzyme, toxin, drug
25 or a prodrug. Therapeutic agents of the present invention also include agonists and antagonists of SSG polypeptides and vaccines capable of inducing an immune response against SSG polypeptides. Such agents can be readily identified in accordance with the teachings herein.

30 Other objects, features, advantages and aspects of the present invention will become apparent to those of skill in the art from the following description. It should be understood, however, that the following description and the specific examples, while indicating preferred embodiments of
35 the invention, are given by way of illustration only. Various

changes and modifications within the spirit and scope of the disclosed invention will become readily apparent to those skilled in the art from reading the following description and from reading the other parts of the present disclosure.

5 GLOSSARY

The following illustrative explanations are provided to facilitate understanding of certain terms used frequently herein. The explanations are provided as a convenience and are not limitative of the invention.

10 DIGESTION of DNA refers to catalytic cleavage of the DNA
with a restriction enzyme that acts only at certain sequences
in the DNA. The various restriction enzymes referred to herein
are commercially available and their reaction conditions,
cofactors and other requirements for use are known and routine
15 to the skilled artisan.

For analytical purposes, typically, 1 μg of plasmid or DNA fragment is digested with about 2 units of enzyme in about 20 μl of reaction buffer. For the purpose of isolating DNA fragments for plasmid construction, typically 5 to 50 μg of DNA are digested with 20 to 250 units of enzyme in proportionately larger volumes.

Appropriate buffers and substrate amounts for particular restriction enzymes are described in standard laboratory manuals, such as those referenced below, and they are
25 specified by commercial suppliers.

Incubation times of about 1 hour at 37°C are ordinarily used, but conditions may vary in accordance with standard procedures, the supplier's instructions and the particulars of the reaction. After digestion, reactions may be analyzed, and fragments may be purified by electrophoresis through an agarose or polyacrylamide gel, using well known methods that are routine for those skilled in the art.

GENETIC ELEMENT generally means a polynucleotide comprising a region that encodes a polypeptide or a region

that regulates transcription or translation or other processes important to expression of the polypeptide in a host cell, or a polynucleotide comprising both a region that encodes a polypeptide and a region operably linked thereto that
5 regulates expression.

Genetic elements may be comprised within a vector that replicates as an episomal element; that is, as a molecule physically independent of the host cell genome. They may be comprised within mini-chromosomes, such as those that arise
10 during amplification of transfected DNA by methotrexate selection in eukaryotic cells. Genetic elements also may be comprised within a host cell genome; not in their natural state but, rather, following manipulation such as isolation, cloning and introduction into a host cell in the form of
15 purified DNA or in a vector, among others.

ISOLATED means altered "by the hand of man" from its natural state; i.e., that, if it occurs in nature, it has been changed or removed from its original environment, or both.

For example, a naturally occurring polynucleotide or a
20 polypeptide naturally present in a living animal in its natural state is not "isolated," but the same polynucleotide or polypeptide separated from the coexisting materials of its natural state is "isolated", as the term is employed herein. For example, with respect to polynucleotides, the term
25 isolated means that it is separated from the chromosome and cell in which it naturally occurs.

As part of or following isolation, such polynucleotides can be joined to other polynucleotides, such as DNAs, for mutagenesis, to form fusion proteins, and for propagation or
30 expression in a host, for instance. The isolated polynucleotides, alone or joined to other polynucleotides such as vectors, can be introduced into host cells, in culture or in whole organisms. When introduced into host cells in culture or in whole organisms, such DNAs still would be isolated, as
35 the term is used herein, because they would not be in their

naturally occurring form or environment. Similarly, the polynucleotides and polypeptides may occur in a composition, such as media formulations, solutions for introduction of polynucleotides or polypeptides, for example, into cells, 5 compositions or solutions for chemical or enzymatic reactions, for instance, which are not naturally occurring compositions, and, therein remain isolated polynucleotides or polypeptides within the meaning of that term as it is employed herein.

LIGATION refers to the process of forming phosphodiester 10 bonds between two or more polynucleotides, which most often are double-stranded DNAs. Techniques for ligation are well known to the art and protocols for ligation are described in standard laboratory manuals and references, such as, for instance, Sambrook et al., MOLECULAR CLONING, A LABORATORY 15 MANUAL, 2nd Ed.; Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. (1989) and Maniatis et al., pg. 146, as cited below.

OLIGONUCLEOTIDE(S) refers to relatively short polynucleotides of about 8 to about 50 nucleobases. Often the 20 term refers to single-stranded deoxyribonucleotides, but it can refer as well to single- or double-stranded ribonucleotides, RNA:DNA hybrids and double-stranded DNAs, among others.

Oligonucleotides, such as single-stranded DNA probe 25 oligonucleotides, often are synthesized by chemical methods, such as those implemented on automated oligonucleotide synthesizers. However, oligonucleotides can be made by a variety of other methods, including *in vitro* recombinant DNA-mediated techniques and by expression of DNAs in cells and 30 organisms.

Initially, chemically synthesized DNAs typically are obtained without a 5' phosphate. The 5' ends of such oligonucleotides are not substrates for phosphodiester bond formation by ligation reactions that employ DNA ligases 35 typically used to form recombinant DNA molecules. Where

ligation of such oligonucleotides is desired, a phosphate can be added by standard techniques, such as those that employ a kinase and ATP.

The 3' end of a chemically synthesized oligonucleotide generally has a free hydroxyl group and, in the presence of a ligase, such as T4 DNA ligase, readily will form a phosphodiester bond with a 5' phosphate of another polynucleotide, such as another oligonucleotide. As is well known, this reaction can be prevented selectively, where desired, by removing the 5' phosphates of the other polynucleotide(s) prior to ligation.

PLASMIDS generally are designated herein by a lower case "p" preceded and/or followed by capital letters and/or numbers, in accordance with standard naming conventions that are familiar to those of skill in the art. Starting plasmids disclosed herein are either commercially available, publicly available on an unrestricted basis, or can be constructed from available plasmids by routine application of well known, published procedures. Many plasmids and other cloning and expression vectors that can be used in accordance with the present invention are well known and readily available to those of skill in the art. Moreover, those of skill readily may construct any number of other plasmids suitable for use in the invention. The properties, construction and use of such plasmids, as well as other vectors, in the present invention will be readily apparent to those of skill from the present disclosure.

POLYNUCLEOTIDE(S) generally refers to any polyribonucleotide or polydeoxribonucleotide, which may be unmodified RNA or DNA or modified RNA or DNA. Thus, for instance, polynucleotides as used herein refers to, among others, single- and double-stranded DNA, DNA that is a mixture of single- and double-stranded regions, single- and double-stranded RNA, and RNA that is mixture of single- and double-stranded regions, hybrid molecules comprising DNA and RNA that

may be single-stranded or, more typically, double-stranded or a mixture of single- and double-stranded regions. In addition, polynucleotide as used herein refers to triple-stranded regions comprising RNA or DNA or both RNA and DNA. The strands
5 in such regions may be from the same molecule or from different molecules. The regions may include all of one or more of the molecules, but more typically involve only a region of some of the molecules. One of the molecules of a triple-helical region often is an oligonucleotide.

10 As used herein, the term polynucleotide includes DNAs or RNAs as described above that contain one or more modified bases. Thus, DNAs or RNAs with backbones modified for stability or for other reasons are "polynucleotides" as that term is intended herein. Moreover, DNAs or RNAs comprising
15 unusual bases, such as inosine, or modified bases, such as tritylated bases, to name just two examples, are polynucleotides as the term is used herein.

It will be appreciated that a great variety of modifications have been made to DNA and RNA that serve many
20 useful purposes known to those of skill in the art. The term polynucleotide as it is employed herein embraces such chemically, enzymatically or metabolically modified forms of polynucleotides, as well as the chemical forms of DNA and RNA characteristic of viruses and cells, including simple and
25 complex cells, inter alia.

POLYPEPTIDES, as used herein, includes all polypeptides as described below. The basic structure of polypeptides is well known and has been described in innumerable textbooks and other publications in the art. In this context, the term is
30 used herein to refer to any peptide or protein comprising two or more amino acids joined to each other in a linear chain by peptide bonds. As used herein, the term refers to both short chains, which also commonly are referred to in the art as peptides, oligopeptides and oligomers, for example, and to
35 longer chains, which generally are referred to in the art as

proteins, of which there are many types. It will be appreciated that polypeptides often contain amino acids other than the 20 amino acids commonly referred to as the 20 naturally occurring amino acids, and that many amino acids, including the terminal amino acids, may be modified in a given polypeptide, either by natural processes, such as processing and other post-translational modifications, or by chemical modification techniques which are well known to the art. Even the common modifications that occur naturally in polypeptides are too numerous to list exhaustively here, but they are well described in basic texts and in more detailed monographs, as well as in voluminous research literature, and they are well known to those of skill in the art.

Known modifications which may be present in polypeptides of the present invention include, to name an illustrative few, acetylation, acylation, ADP-ribosylation, amidation, covalent attachment of flavin, covalent attachment of a heme moiety, covalent attachment of a nucleotide or nucleotide derivative, covalent attachment of a lipid or lipid derivative, covalent attachment of phosphatidylinositol, cross-linking, cyclization, disulfide bond formation, demethylation, formation of covalent cross-links, formation of cystine, formation of pyroglutamate, formylation, gamma-carboxylation, glycosylation, GPI anchor formation, hydroxylation, iodination, methylation, myristoylation, oxidation, proteolytic processing, phosphorylation, prenylation, racemization, selenoylation, sulfation, transfer-RNA mediated addition of amino acids to proteins such as arginylation, and ubiquitination.

Such modifications are well known to those of skill and have been described in great detail in the scientific literature. Several particularly common modifications, glycosylation, lipid attachment, sulfation, gamma-carboxylation of glutamic acid residues, hydroxylation and ADP-ribosylation, for instance, are described in most basic

texts, such as, for instance PROTEINS STRUCTURE AND MOLECULAR
PROPERTIES, 2nd Ed., T. E. Creighton, W. H. Freeman and
Company, New York (1993). Many detailed reviews are available
on this subject, such as, for example, those provided by Wold,
5 F., Posttranslational Protein Modifications: Perspectives and
Prospects, pgs. 1-12 in POSTTRANSLATIONAL COVALENT
MODIFICATION OF PROTEINS, B. C. Johnson, Ed., Academic Press,
New York (1983); Seifter et al., Analysis for protein
modifications and nonprotein cofactors, Meth. Enzymol. 182:
10 626-646 (1990) and Rattan et al., Protein Synthesis:
Posttranslational Modifications and Aging, Ann. N.Y. Acad.
Sci. 663: 48-62 (1992).

It will be appreciated, as is well known and as noted
above, that polypeptides are not always entirely linear. For
15 instance, polypeptides may be branched as a result of
ubiquitination, and they may be circular, with or without
branching, generally as a result of posttranslation events,
including natural processing events and events brought about
by human manipulation which do not occur naturally. Circular,
20 branched and branched circular polypeptides may be synthesized
by non-translation natural process and by entirely synthetic
methods, as well.

Modifications can occur anywhere in a polypeptide,
including the peptide backbone, the amino acid side-chains and
25 the amino or carboxyl termini. In fact, blockage of the amino
or carboxyl group in a polypeptide, or both, by a covalent
modification, is common in naturally occurring and synthetic
polypeptides and such modifications may be present in
polypeptides of the present invention, as well. For instance,
30 the amino terminal residue of polypeptides made in *E. coli*,
prior to proteolytic processing, almost invariably will be N-
formylmethionine.

The modifications that occur in a polypeptide often will
be a function of how it is made. For polypeptides made by
35 expressing a cloned gene in a host, for instance, the nature

and extent of the modifications in large part will be determined by the host cell posttranslational modification capacity and the modification signals present in the polypeptide amino acid sequence. For instance, as is well known, glycosylation often does not occur in bacterial hosts such as *E. coli*. Accordingly, when glycosylation is desired, a polypeptide should be expressed in a glycosylating host, generally a eukaryotic cell. Insect cells often carry out the same posttranslational glycosylations as mammalian cells and, for this reason, insect cell expression systems have been developed to express efficiently mammalian proteins having native patterns of glycosylation, *inter alia*. Similar considerations apply to other modifications.

It will be appreciated that the same type of modification may be present in the same or varying degree at several sites in a given polypeptide. Also, a given polypeptide may contain many types of modifications.

In general, as used herein, the term polypeptide encompasses all such modifications, particularly those that are present in polypeptides synthesized by expressing a polynucleotide in a host cell.

VARIANT(S) of polynucleotides or polypeptides, as the term is used herein, are polynucleotides or polypeptides that differ from a reference polynucleotide or polypeptide, respectively. Variants in this sense are described below and elsewhere in the present disclosure in greater detail.

A variant may comprise a polynucleotide that differs in nucleotide sequence from another, reference polynucleotide. Generally, differences are limited so that the nucleotide sequences of the reference and the variant are closely similar overall and, in many regions, identical.

As noted below, changes in the nucleotide sequence of the variant may be silent. That is, they may not alter the amino acids encoded by the polynucleotide. Where alterations are limited to silent changes of this type a variant will encode

a polypeptide with the same amino acid sequence as the reference. Also as noted below, changes in the nucleotide sequence of the variant may alter the amino acid sequence of a polypeptide encoded by the reference polynucleotide. Such
5 nucleotide changes may result in amino acid substitutions, additions, deletions, fusions and truncations in the polypeptide encoded by the reference sequence, as discussed below.

A variant may also comprise a polypeptide that differs
10 in amino acid sequence from another, reference polypeptide. Generally, differences are limited so that the sequences of the reference and the variant are closely similar overall and, in many regions, identical.

A variant and reference polypeptide may differ in amino
15 acid sequence by one or more substitutions, additions, deletions, fusions and truncations, which may be present in any combination.

RECEPTOR MOLECULE, as used herein, refers to molecules which bind or interact specifically with SSG polypeptides of the present invention and is inclusive not only of classic
20 receptors, which are preferred, but also other molecules that specifically bind to or interact with polypeptides of the invention (which also may be referred to as "binding molecules" and "interaction molecules," respectively and as
25 "SSG binding or interaction molecules". Binding between polypeptides of the invention and such molecules, including receptor or binding or interaction molecules may be exclusive to polypeptides of the invention, which is very highly preferred, or it may be highly specific for polypeptides of
30 the invention, which is highly preferred, or it may be highly specific to a group of proteins that includes polypeptides of the invention, which is preferred, or it may be specific to several groups of proteins at least one of which includes polypeptides of the invention.

Receptors also may be non-naturally occurring, such as antibodies and antibody-derived reagents that bind to polypeptides of the invention.

5 DETAILED DESCRIPTION OF THE INVENTION

Polynucleotides and polypeptides

The present invention relates to newly identified isolated polynucleotides and polypeptides encoded thereby which are upregulated in or specific to stomach cancer tissue.

10 These polynucleotides and the polypeptides encoded thereby are believed to be useful as diagnostic markers for cancer, and in particular stomach cancer.

15 For purposes of the present invention, by polynucleotides it is meant to include isolated nucleic acid sequences comprising single- and double-stranded DNA, DNA that is a mixture of single- and double-stranded regions, single and double-stranded RNA or hybrids thereof wherein the sequences comprise SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 or 13, fragments of at least 15 contiguous nucleobases of SEQ
20 ID NO: 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 or 13, nucleic acid sequences which, due to degeneracy in genetic coding, comprise variations in polynucleotide sequence as compared to SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 or 13, but which still encode the same protein, and nucleic acid
25 sequences which are capable of hybridizing under stringent conditions to the antisense sequence of SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 or 13. By stringent conditions it is meant that hybridization will occur only if there is at least 95%, and more preferably at least 97% identity between
30 the sequences. RNA sequences may be in the form of mRNA while DNA sequences may be in the form of cDNA or genomic DNA obtained by cloning or produced by chemical synthetic techniques or by a combination thereof. As used herein, the term polynucleotide also includes DNAs or RNAs, as described
35 above, that contain one or more modified bases. Examples of

modified bases include, but are not limited to, backbone modifications to increase stability and incorporation of unusual bases such as inosine or tritylated bases.

For purposes of the present invention, by polypeptides
5 it is meant to include the recombinant, natural and synthetic polypeptides with amino acid sequences encoded by the polynucleotides of SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 or 13, or fragments or variants thereof with similar activities and/or levels in cancerous tissues to the amino
10 acid sequences encoded by the polynucleotides of the present invention. Among preferred variants are those that vary from the polypeptides encoded by SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 or 13 by conservative amino acid substitutions. Conservative amino acid substitutions
15 typically include replacement, one for another, of the aliphatic amino acids such as Ala, Val, Leu and Ile, the hydroxyl residues Ser and Thr, the acidic residues Asp and Glu, the amide residues Asn and Gln, the basic residues Lys and Arg, and the aromatic residues Phe and Tyr.

20 Using suppression subtractive hybridization, it has now been found that these polynucleotides, and the polypeptides encoded thereby, are upregulated in, or specific to, stomach cancer tissue. Thus, it is believed that these polynucleotides and polypeptides, also referred to herein as
25 Stomach Cancer Specific Genes or SSGs, are useful as diagnostic markers for stomach cancer, as well as otherwise described herein.

Fragments

Also among preferred embodiments of this aspect of the
30 present invention are polypeptides comprising fragments of SSGs, and fragments of variants and derivatives of the SSGs.

In this regard a fragment is a polypeptide having an amino acid sequence that entirely is the same as part but not all of the amino acid sequences encoded by the SSGs of SEQ ID

NO: 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, or 13 and variants or derivatives thereof.

Such fragments may be "free-standing," i.e., not part of or fused to other amino acids or polypeptides, or they may be
5 comprised within a larger polypeptide of which they form a part or region. When comprised within a larger polypeptide, the presently discussed fragments most preferably form a single continuous region. However, several fragments may be comprised within a single larger polypeptide. For instance,
10 certain preferred embodiments relate to a fragment of a SSG polypeptide of the present invention comprised within a precursor polypeptide designed for expression in a host and having heterologous pre- and pro-polypeptide regions fused to the amino terminus of the SSG fragment and an additional
15 region fused to the carboxyl terminus of the fragment. Therefore, fragments in one aspect of the meaning intended herein, refers to the portion or portions of a fusion polypeptide or fusion protein derived from a SSG of the present invention.

20 As representative examples of polypeptide fragments of the invention, there may be mentioned those which have from about 15 to about 139 amino acids.

In this context "about" includes the particularly recited range and ranges larger or smaller by several, a few, 5, 4,
25 3, 2 or 1 amino acid at either extreme or at both extremes. Highly preferred in this regard are the recited ranges plus or minus as many as 5 amino acids at either or at both extremes. Particularly highly preferred are the recited ranges plus or minus as many as 3 amino acids at either or at
30 both the recited extremes. Especially preferred are ranges plus or minus 1 amino acid at either or at both extremes or the recited ranges with no additions or deletions. Most highly preferred of all in this regard are fragments from about 15 to about 45 amino acids.

Among especially preferred fragments of the invention are truncation mutants of the SSGs. Truncation mutants include SSG polypeptides encoded by SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, or 13 or variants or derivatives thereof, except for deletion of a continuous series of residues (that is, a continuous region, part or portion) that includes the amino terminus, or a continuous series of residues that includes the carboxyl terminus or, as in double truncation mutants, deletion of two continuous series of residues, one including the amino terminus and one including the carboxyl terminus. Fragments having the size ranges set out above also are preferred embodiments of truncation fragments, which are especially preferred among fragments generally.

Also preferred in this aspect of the invention are fragments characterized by structural or functional attributes of the SSGs. Preferred embodiments of the invention in this regard include fragments that comprise alpha-helix and alpha-helix forming regions ("alpha-regions"), beta-sheet and beta-sheet-forming regions ("beta-regions"), turn and turn-forming regions ("turn-regions"), coil and coil-forming regions ("coil-regions"), hydrophilic regions, hydrophobic regions, alpha amphipathic regions, beta amphipathic regions, flexible regions, surface-forming regions and high antigenic index regions of SSGs. Garnier-Robson alpha-regions, beta-regions, turn-regions and coil-regions, Chou-Fasman alpha-regions, beta-regions and turn-regions, Kyte-Doolittle hydrophilic regions and hydrophilic regions, Eisenberg alpha and beta amphipathic regions, Karplus-Schulz flexible regions, Emini surface-forming regions and Jameson-Wolf high antigenic index regions are particularly preferred. Among highly preferred fragments in this regard are those that comprise regions of SSGs that combine several structural features, such as several of the features set out above. Such regions may be comprised within a larger polypeptide or may be by themselves a preferred fragment of the present invention, as discussed

above. It will be appreciated that the term "about" as used in this paragraph has the meaning set out above regarding fragments in general.

Further preferred regions are those that mediate activities of SSGs. Most highly preferred in this regard are fragments that have a chemical, biological or other activity of a SSG, including those with a similar activity or an improved activity, or with a decreased undesirable activity. Highly preferred in this regard are fragments that contain regions that are homologs in sequence, or in position, or in both sequence and to active regions of related polypeptides, and which include lung specific-binding proteins. Among particularly preferred fragments in these regards are truncation mutants, as discussed above.

It will be appreciated that the invention also relates to, among others, polynucleotides encoding the aforementioned fragments, polynucleotides that hybridize to polynucleotides encoding the fragments, particularly those that hybridize under stringent conditions, and polynucleotides, such as PCR primers, for amplifying polynucleotides that encode the fragments. In these regards, preferred polynucleotides are those that correspondent to the preferred fragments, as discussed above.

Vectors, host cells, expression

The present invention also relates to vectors which include polynucleotides of the present invention, host cells which are genetically engineered with vectors of the invention and the production of polypeptides of the invention by recombinant techniques.

Host cells can be genetically engineered to incorporate polynucleotides and express polypeptides of the present invention. For instance, polynucleotides may be introduced into host cells using well known techniques of infection, transduction, transfection, transvection and transformation. The polynucleotides may be introduced alone or with other

polynucleotides. Such other polynucleotides may be introduced independently, co-introduced or introduced joined to the polynucleotides of the invention.

Thus, for instance, polynucleotides of the invention may
5 be transfected into host cells with another, separate, polynucleotide encoding a selectable marker, using standard techniques for co-transfection and selection in, for instance, mammalian cells. In this case the polynucleotides generally will be stably incorporated into the host cell genome.

10 Alternatively, the polynucleotides may be joined to a vector containing a selectable marker for propagation in a host. The vector construct may be introduced into host cells by the aforementioned techniques. Generally, a plasmid vector is introduced as DNA in a precipitate, such as a calcium
15 phosphate precipitate, or in a complex with a charged lipid. Electroporation also may be used to introduce polynucleotides into a host. If the vector is a virus, it may be packaged *in vitro* or introduced into a packaging cell and the packaged virus may be transduced into cells. A wide variety of
20 techniques suitable for making polynucleotides and for introducing polynucleotides into cells in accordance with this aspect of the invention are well known and routine to those of skill in the art. Such techniques are reviewed at length in Sambrook et al. cited above, which is illustrative of the
25 many laboratory manuals that detail these techniques. In accordance with this aspect of the invention the vector may be, for example, a plasmid vector, a single- or double-stranded phage vector, or a single- or double-stranded RNA or DNA viral vector. Such vectors may be introduced into cells
30 as polynucleotides, preferably DNA, by well known techniques for introducing DNA and RNA into cells. The vectors, in the case of phage and viral vectors also may be and preferably are introduced into cells as packaged or encapsidated virus by well known techniques for infection and transduction. Viral
35 vectors may be replication competent or replication defective.

In the latter case viral propagation generally will occur only in complementing host cells.

Preferred among vectors, in certain respects, are those for expression of polynucleotides and polypeptides of the present invention. Generally, such vectors comprise cis-acting control regions effective for expression in a host operatively linked to the polynucleotide to be expressed. Appropriate trans-acting factors either are supplied by the host, supplied by a complementing vector or supplied by the vector itself upon introduction into the host.

In certain preferred embodiments in this regard, the vectors provide for specific expression. Such specific expression may be inducible expression or expression only in certain types of cells or both inducible and cell-specific. Particularly preferred among inducible vectors are vectors that can be induced for expression by environmental factors that are easy to manipulate, such as temperature and nutrient additives. A variety of vectors suitable to this aspect of the invention, including constitutive and inducible expression vectors for use in prokaryotic and eukaryotic hosts, are well known and employed routinely by those of skill in the art.

The engineered host cells can be cultured in conventional nutrient media, which may be modified as appropriate for, inter alia, activating promoters, selecting transformants or amplifying genes. Culture conditions, such as temperature, pH and the like, previously used with the host cell selected for expression generally will be suitable for expression of polypeptides of the present invention as will be apparent to those of skill in the art.

A great variety of expression vectors can be used to express a polypeptide of the invention. Such vectors include chromosomal, episomal and virus-derived vectors e.g., vectors derived from bacterial plasmids, from bacteriophage, from yeast episomes, from yeast chromosomal elements, from viruses such as baculoviruses, papova viruses, such as SV40, vaccinia

viruses, adenoviruses, fowl pox viruses, pseudorabies viruses and retroviruses, and vectors derived from combinations thereof, such as those derived from plasmid and bacteriophage genetic elements, such as cosmids and phagemids, all may be
5 used for expression in accordance with this aspect of the present invention. Generally, any vector suitable to maintain, propagate or express polynucleotides to express a polypeptide in a host may be used for expression in this regard.

The appropriate DNA sequence may be inserted into the
10 vector by any of a variety of well-known and routine techniques. In general, a DNA sequence for expression is joined to an expression vector by cleaving the DNA sequence and the expression vector with one or more restriction endonucleases and then joining the restriction fragments
15 together using T4 DNA ligase. Procedures for restriction and ligation that can be used to this end are well known and routine to those of skill. Suitable procedures in this regard, and for constructing expression vectors using alternative techniques, which also are well known and routine to those
20 skill, are set forth in great detail in Sambrook et al. cited elsewhere herein.

The DNA sequence in the expression vector is operatively linked to appropriate expression control sequence(s), including, for instance, a promoter to direct mRNA
25 transcription. Representatives of such promoters include the phage lambda PL promoter, the *E. coli* lac, trp and tac promoters, the SV40 early and late promoters and promoters of retroviral LTRs, to name just a few of the well-known promoters. It will be understood that numerous promoters not
30 mentioned are suitable for use in this aspect of the invention are well known and readily may be employed by those of skill in the manner illustrated by the discussion and the examples herein.

In general, expression constructs will contain sites for
35 transcription initiation and termination, and, in the

transcribed region, a ribosome binding site for translation. The coding portion of the mature transcripts expressed by the constructs will include a translation initiating AUG at the beginning and a termination codon appropriately positioned at the end of the polypeptide to be translated.

In addition, the constructs may contain control regions that regulate as well as engender expression. Generally, in accordance with many commonly practiced procedures, such regions will operate by controlling transcription, such as repressor binding sites and enhancers, among others.

Vectors for propagation and expression generally will include selectable markers. Such markers also may be suitable for amplification or the vectors may contain additional markers for this purpose. In this regard, the expression vectors preferably contain one or more selectable marker genes to provide a phenotypic trait for selection of transformed host cells. Preferred markers include dihydrofolate reductase or neomycin resistance for eukaryotic cell culture, and tetracycline or ampicillin resistance genes for culturing *E. coli* and other bacteria.

The vector containing the appropriate DNA sequence as described elsewhere herein, as well as an appropriate promoter, and other appropriate control sequences, may be introduced into an appropriate host using a variety of well known techniques suitable to expression therein of a desired polypeptide. Representative examples of appropriate hosts include bacterial cells, such as *E. coli*, *Streptomyces* and *Salmonella typhimurium* cells; fungal cells, such as yeast cells; insect cells such as *Drosophila* S2 and *Spodoptera* Sf9 cells; animal cells such as CHO, COS and Bowes melanoma cells; and plant cells. Hosts for a great variety of expression constructs are well known, and those of skill will be enabled by the present disclosure readily to select a host for expressing a polypeptides in accordance with this aspect of the present invention.

More particularly, the present invention also includes recombinant constructs, such as expression constructs, comprising one or more of the sequences described above. The constructs comprise a vector, such as a plasmid or viral
5 vector, into which such a sequence of the invention has been inserted. The sequence may be inserted in a forward or reverse orientation. In certain preferred embodiments in this regard, the construct further comprises regulatory sequences, including, for example, a promoter, operably linked to the
10 sequence. Large numbers of suitable vectors and promoters are known to those of skill in the art, and there are many commercially available vectors suitable for use in the present invention.

The following vectors, which are commercially available,
15 are provided by way of example. Among vectors preferred for use in bacteria are pQE70, pQE60 and pQE-9, available from Qiagen; pBS vectors, Phagescript vectors, Bluescript vectors, pNH8A, pNH16a, pNH18A, pNH46A, available from Stratagene; and ptrc99a, pKK223-3, pKK233-3, pDR540, pRIT5 available from
20 Pharmacia. Among preferred eukaryotic vectors are PWLNEO, pSV2CAT, pOG44, pXT1 and pSG available from Stratagene; and pSVK3, pBPV, pMSG and pSVL available from Pharmacia. These vectors are listed solely by way of illustration of the many commercially available and well known vectors that are
25 available to those of skill in the art for use in accordance with this aspect of the present invention. It will be appreciated that any other plasmid or vector suitable for, for example, introduction, maintenance, propagation or expression of a polynucleotide or polypeptide of the invention in a host
30 may be used in this aspect of the invention.

Promoter regions can be selected from any desired gene using vectors that contain a reporter transcription unit lacking a promoter region, such as a chloramphenicol acetyl transferase ("cat") transcription unit, downstream of
35 restriction site or sites for introducing a candidate promoter

fragment; i.e., a fragment that may contain a promoter. As is well known, introduction into the vector of a promoter-containing fragment at the restriction site upstream of the cat gene engenders production of CAT activity, which can be
5 detected by standard CAT assays. Vectors suitable to this end are well known and readily available. Two such vectors are pKK232-8 and pCM7. Thus, promoters for expression of polynucleotides of the present invention include not only well known and readily available promoters, but also promoters that
10 readily may be obtained by the foregoing technique, using a reporter gene.

Among known bacterial promoters suitable for expression of polynucleotides and polypeptides in accordance with the present invention are the *E. coli* lacI and lacZ and promoters,
15 the T3 and T7 promoters, the gpt promoter, the lambda PR, PL promoters and the trp promoter. Among known eukaryotic promoters suitable in this regard are the CMV immediate early promoter, the HSV thymidine kinase promoter, the early and late SV40 promoters, the promoters of retroviral LTRs, such
20 as those of the Rous sarcoma virus ("RSV"), and metallothionein promoters, such as the mouse metallothionein-I promoter.

Selection of appropriate vectors and promoters for expression in a host cell is a well known procedure and the
25 requisite techniques for expression vector construction, introduction of the vector into the host and expression in the host are routine skills in the art.

The present invention also relates to host cells containing the above-described constructs discussed above. The
30 host cell can be a higher eukaryotic cell, such as a mammalian cell, or a lower eukaryotic cell, such as a yeast cell, or the host cell can be a prokaryotic cell, such as a bacterial cell.

Introduction of the construct into the host cell can be effected by calcium phosphate transfection, DEAE-dextran
35 mediated transfection, cationic lipid-mediated transfection,

electroporation, transduction, infection or other methods. Such methods are described in many standard laboratory manuals, such as Davis et al. BASIC METHODS IN MOLECULAR BIOLOGY, (1986).

5 Constructs in host cells can be used in a conventional manner to produce the gene product encoded by the recombinant sequence. Alternatively, the polypeptides of the invention can be synthetically produced by conventional peptide synthesizers.

10 Mature proteins can be expressed in mammalian cells, yeast, bacteria, or other cells under the control of appropriate promoters. Cell-free translation systems can also be employed to produce such proteins using RNAs derived from the DNA constructs of the present invention. Appropriate
15 cloning and expression vectors for use with prokaryotic and eukaryotic hosts are described by Sambrook et al., MOLECULAR CLONING: A LABORATORY MANUAL, 2nd Ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. (1989).

20 Generally, recombinant expression vectors will include origins of replication, a promoter derived from a highly-expressed gene to direct transcription of a downstream structural sequence, and a selectable marker to permit isolation of vector containing cells after exposure to the vector. Among suitable promoters are those derived from the
25 genes that encode glycolytic enzymes such as 3-phosphoglycerate kinase ("PGK"), a-factor, acid phosphatase, and heat shock proteins, among others. Selectable markers include the ampicillin resistance gene of *E. coli* and the *trp1* gene of *S. cerevisiae*.

30 Transcription of the DNA encoding the polypeptides of the present invention by higher eukaryotes may be increased by inserting an enhancer sequence into the vector. Enhancers are cis-acting elements of DNA, usually about from 10 to 300 bp that act to increase transcriptional activity of a promoter
35 in a given host cell-type. Examples of enhancers include the

SV40 enhancer, which is located on the late side of the replication origin at bp 100 to 270, the cytomegalovirus early promoter enhancer, the polyoma enhancer on the late side of the replication origin, and adenovirus enhancers.

5 Polynucleotides of the invention, encoding the heterologous structural sequence of a polypeptide of the invention generally will be inserted into the vector using standard techniques so that it is operably linked to the promoter for expression. The polynucleotide will be positioned
10 so that the transcription start site is located appropriately 5' to a ribosome binding site. The ribosome binding site will be 5' to the AUG that initiates translation of the polypeptide to be expressed. Generally, there will be no other open reading frames that begin with an initiation codon, usually
15 AUG, and lie between the ribosome binding site and the initiating AUG. Also, generally, there will be a translation stop codon at the end of the polypeptide and there will be a polyadenylation signal and a transcription termination signal appropriately disposed at the 3' end of the transcribed
20 region.

For secretion of the translated protein into the lumen of the endoplasmic reticulum, into the periplasmic space or into the extracellular environment, appropriate secretion signals may be incorporated into the expressed polypeptide.
25 The signals may be endogenous to the polypeptide or they may be heterologous signals.

The polypeptide may be expressed in a modified form, such as a fusion protein, and may include not only secretion signals but also additional heterologous functional regions.
30 Thus, for instance, a region of additional amino acids, particularly charged amino acids, may be added to the N-terminus of the polypeptide to improve stability and persistence in the host cell, during purification or during subsequent handling and storage. A region also may be added
35 to the polypeptide to facilitate purification. Such regions

may be removed prior to final preparation of the polypeptide. The addition of peptide moieties to polypeptides to engender secretion or excretion, to improve stability and to facilitate purification, among others, are familiar and routine techniques in the art.

Suitable prokaryotic hosts for propagation, maintenance or expression of polynucleotides and polypeptides in accordance with the invention include *Escherichia coli*, *Bacillus subtilis* and *Salmonella typhimurium*. Various species of *Pseudomonas*, *Streptomyces*, and *Staphylococcus* are suitable hosts in this regard. Moreover, many other hosts also known to those of skill may be employed in this regard.

As a representative but non-limiting example, useful expression vectors for bacterial use can comprise a selectable marker and bacterial origin of replication derived from commercially available plasmids comprising genetic elements of the well known cloning vector pBR322. Such commercial vectors include, for example, pKK223-3 (Pharmacia Fine Chemicals, Uppsala, Sweden) and GEM1 (Promega Biotec, Madison, Wis., USA). These pBR322 "backbone" sections are combined with an appropriate promoter and the structural sequence to be expressed.

Following transformation of a suitable host strain and growth of the host strain to an appropriate cell density, where the selected promoter is inducible it is induced by appropriate means (e.g., temperature shift or exposure to chemical inducer) and cells are cultured for an additional period.

Cells typically then are harvested by centrifugation, disrupted by physical or chemical means, and the resulting crude extract retained for further purification. Microbial cells employed in expression of proteins can be disrupted by any convenient method, including freeze-thaw cycling, sonication, mechanical disruption, or use of cell lysing

agents, such methods are well know to those skilled in the art.

Various mammalian cell culture systems can be employed for expression, as well. Examples of mammalian expression systems include the COS-7 lines of monkey kidney fibroblast, described in Gluzman et al., Cell 23: 175 (1981). Other cell lines capable of expressing a compatible vector include for example, the C127, 3T3, CHO, HeLa, human kidney 293 and BHK cell lines.

10 Mammalian expression vectors will comprise an origin of replication, a suitable promoter and enhancer, and also any necessary ribosome binding sites, polyadenylation sites, splice donor and acceptor sites, transcriptional termination sequences, and 5' flanking non-transcribed sequences that are
15 necessary for expression. In certain preferred embodiments in this regard DNA sequences derived from the SV40 splice sites, and the SV40 polyadenylation sites are used for required non-transcribed genetic elements of these types.

The SSG polypeptide can be recovered and purified from
20 recombinant cell cultures by well-known methods including ammonium sulfate or ethanol precipitation, acid extraction, anion or cation exchange chromatography, phosphocellulose chromatography, hydrophobic interaction chromatography, affinity chromatography, hydroxylapatite chromatography and
25 lectin chromatography. Most preferably, high performance liquid chromatography ("HPLC") is employed for purification. Well known techniques for refolding protein may be employed to regenerate active conformation when the polypeptide is denatured during isolation and or purification.

30 Polypeptides of the present invention include naturally purified products, products of chemical synthetic procedures, and products produced by recombinant techniques from a prokaryotic or eukaryotic host, including, for example, bacterial, yeast, higher plant, insect and mammalian cells.
35 Depending upon the host employed in a recombinant production

procedure, the polypeptides of the present invention may be glycosylated or may be non-glycosylated. In addition, polypeptides of the invention may also include an initial modified methionine residue, in some cases as a result of
5 host-mediated processes.

SSG polynucleotides and polypeptides may be used in accordance with the present invention for a variety of applications, particularly those that make use of the chemical and biological properties of the SSGs. Additional
10 applications relate to diagnosis and to treatment of disorders of cells, tissues and organisms. These aspects of the invention are illustrated further by the following discussion.

Antibodies

The SSG polypeptides of the invention or their fragments
15 or variants thereof, or cells expressing them can be used as immunogens to produce antibodies immunospecific for the SSG polypeptides. The term "immunospecific" means that the antibodies have substantially greater affinity for the polypeptides of the invention than their affinity for other
20 related polypeptides in the prior art. These antibodies can be polyclonal or monoclonal. In addition, by the term "antibody", it is meant to include chimeric, single chain and humanized and fully human antibodies as well as Fab fragments or products of Fab expression libraries.

25 Antibodies generated against the SSG polypeptides can be obtained by administering the polypeptides or epitope-bearing fragments, variants or cells to an animal, preferably a nonhuman, using routine protocols. For preparation of monoclonal antibodies, any technique which provides antibodies
30 produced by continuous cell line cultures can be used. Examples include the hybridoma technique (Kohler, G. and Milstein, C., *Nature* (1975) 256:495-497), the trioma technique, the human B-cell hybridoma technique (Kozbor et al., *Immunology Today* (1983) 4:72) and the EBV-hybridoma

technique (Cole et al., MONOCLONAL ANTIBODIES AND CANCER THERAPY, pp. 77-96, Alan R. Liss, Inc., 1985).

Techniques for the production of single chain antibodies (U.S. Patent No. 4,946,778) can also be adapted to produce
5 single chain antibodies to polypeptides of this invention. Also, transgenic mice, or other organisms including other mammals, can be used to express humanized antibodies.

The above-described antibodies can be used to isolate or to identify clones expressing the polypeptide or to purify the
10 polypeptides by affinity chromatography.

Antibodies against SSG polypeptides can also be used to treat stomach cancer, among others.

Diagnostic Tools

The present invention also relates to diagnostic tools
15 such as antibodies which are immunospecific for SSGs or labeled oligonucleotide probes which hybridize to SSGs.

Antibodies immunospecific for SSGs are described in detail in the preceding section.

Antisense oligonucleotides which hybridize to a portion
20 of a polynucleotide of the present invention can be chemically synthesized via an automated oligonucleotide synthesizer or produced via alternative methods such as *in vitro* recombinant DNA-mediated techniques and by expression of DNA in cells and organisms. By the term "oligonucleotide" it is meant
25 relatively short polynucleotides of about 8 to 50 nucleobases. Most often oligonucleotides comprise single-stranded deoxyribonucleotides. However, oligonucleotides may also comprise single-or double-stranded ribonucleotide, RNA:DNA hybrids and double-stranded DNAs.

30 Methods of Use

The present invention also relates to assays and methods, both quantitative and qualitative, for detecting, diagnosing, monitoring, staging and prognosticating cancers by comparing levels of SSG in a human patient with those of SSG in a normal
35 human control. For purposes of the present invention, what

is meant by "SSG levels" is, among other things, native protein expressed by a polynucleotide sequence comprising SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 or 13. By SSG it is also meant herein polynucleotides which, due to degeneracy in genetic coding, comprise variations in nucleotide sequence as compared to SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 or 13, but which still encode the same protein. The native protein being detected may be whole, a breakdown product, a complex of molecules or chemically modified. In the alternative, what is meant by SSG as used herein, means the native mRNA encoded by the gene comprising the polynucleotide sequence of SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 or 13, or a polynucleotide which is capable of hybridizing under stringent conditions to the antisense sequence of SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 or 13. Such levels are preferably determined in at least one of cells, tissues and/or bodily fluids, including determination of normal and abnormal levels. Thus, for instance, a diagnostic assay in accordance with the invention for diagnosing overexpression of SSG protein compared to normal control bodily fluids, cells, or tissue samples may be used to diagnose the presence of stomach cancer.

All the methods of the present invention may optionally include determining the levels of other cancer markers as well as SSG. Other cancer markers, in addition to SSG, useful in the present invention will depend on the cancer being tested and are known to those of skill in the art.

Diagnostic Assays

The present invention provides methods for diagnosing the presence of cancer, and in particular stomach cancer, by analyzing for changes in levels of SSG in cells, tissues or bodily fluids compared with levels of SSG in cells, tissues or bodily fluids of preferably the same type from a normal human control, wherein an increase in levels of SSG in the

patient versus the normal human control is associated with the presence of stomach cancer.

Without limiting the instant invention, typically, for a quantitative diagnostic assay a positive result indicating
5 the patient being tested has cancer is one in which cells, tissues or bodily fluid levels of the cancer marker, such as SSG, are at least two times higher, and most preferably are at least five times higher, than in preferably the same cells, tissues or bodily fluid of a normal human control.

10 The present invention also provides a method of diagnosing metastatic stomach cancer in a patient having stomach cancer which has not yet metastasized for the onset of metastasis. In the method of the present invention, a human cancer patient suspected of having stomach cancer which
15 may have metastasized (but which was not previously known to have metastasized) is identified. This is accomplished by a variety of means known to those of skill in the art.

In the present invention, determining the presence of SSG levels in cells, tissues or bodily fluid, is particularly
20 useful for discriminating between stomach cancer which has not metastasized and stomach cancer which has metastasized. Existing techniques have difficulty discriminating between stomach cancer which has metastasized and stomach cancer which has not metastasized and proper treatment selection is often
25 dependent upon such knowledge.

In the present invention, the cancer marker levels are measured in such cells, tissues or bodily fluid is SSG, and are then compared with levels of SSG in preferably the same cells, tissue or bodily fluid type of a normal human control.
30 That is, if the cancer marker being observed is SSG in serum, this level is preferably compared with the level of SSG in serum of a normal human control. An increase in the SSG in the patient versus the normal human control is associated with stomach cancer which has metastasized.

Without limiting the instant invention, typically, for a quantitative diagnostic assay a positive result indicating the cancer in the patient being tested or monitored has metastasized is one in which cells, tissues or bodily fluid levels of the cancer marker, such as SSG, are at least two times higher, and most preferably are at least five times higher, than in preferably the same cells, tissues or bodily fluid of a normal patient.

Normal human control as used herein includes a human patient without cancer and/or non cancerous samples from the patient. In the methods for diagnosing or monitoring for metastasis, normal human control may preferably also include samples from a human patient that is determined by reliable methods to have stomach cancer which has not metastasized.

15 **Staging**

The invention also provides a method of staging stomach cancer in a human patient. The method comprises identifying a human patient having such cancer and analyzing cells, tissues or bodily fluid from such human patient for SSG. The SSG levels determined in the patient are then compared with levels of SSG in preferably the same cells, tissues or bodily fluid type of a normal human control, wherein an increase in SSG levels in the human patient versus the normal human control is associated with a cancer which is progressing and a decrease in the levels of SSG (but still increased over true normal levels) is associated with a cancer which is regressing or in remission.

Monitoring

Further provided is a method of monitoring stomach cancer in a human patient having such cancer for the onset of metastasis. The method comprises identifying a human patient having such cancer that is not known to have metastasized; periodically analyzing cells, tissues or bodily fluid from such human patient for SSG; and comparing the SSG levels determined in the human patient with levels of SSG in

preferably the same cells, tissues or bodily fluid type of a normal human control, wherein an increase in SSG levels in the human patient versus the normal human control is associated with a cancer which has metastasized. In this method, normal human control samples may also include prior patient samples.

Further provided by this invention is a method of monitoring the change in stage of stomach cancer in a human patient having such cancer. The method comprises identifying a human patient having such cancer; periodically analyzing cells, tissues or bodily fluid from such human patient for SSG; and comparing the SSG levels determined in the human patient with levels of SSG in preferably the same cells, tissues or bodily fluid type of a normal human control, wherein an increase in SSG levels in the human patient versus the normal human control is associated with a cancer which is progressing in stage and a decrease in the levels of SSG is associated with a cancer which is regressing in stage or in remission. In this method, normal human control samples may also include prior patient samples.

Monitoring a patient for onset of metastasis is periodic and preferably done on a quarterly basis. However, this may be done more or less frequently depending on the cancer, the particular patient, and the stage of the cancer.

Prognostic Testing and Clinical Trial Monitoring

The methods described herein can further be utilized as prognostic assays to identify subjects having or at risk of developing a disease or disorder associated with increased levels of SSG. The present invention provides a method in which a test sample is obtained from a human patient and SSG is detected. The presence of higher SSG levels as compared to normal human controls is diagnostic for the human patient being at risk for developing cancer, particularly stomach cancer.

The effectiveness of therapeutic agents to decrease expression or activity of the SSGs of the invention can also

be monitored by analyzing levels of expression of the SSGs in a human patient in clinical trials or in *in vitro* screening assays such as in human cells. In this way, the gene expression pattern can serve as a marker, indicative of the physiological response of the human patient, or cells as the case may be, to the agent being tested.

Detection of genetic lesions or mutations

The methods of the present invention can also be used to detect genetic lesions or mutations in SSG, thereby determining if a human with the genetic lesion is at risk for stomach cancer or has stomach cancer. Genetic lesions can be detected, for example, by ascertaining the existence of a deletion and/or addition and/or substitution of one or more nucleotides from the SSGs of this invention, a chromosomal rearrangement of SSG, aberrant modification of SSG (such as of the methylation pattern of the genomic DNA), the presence of a non-wild type splicing pattern of a mRNA transcript of SSG, allelic loss of SSG, and/or inappropriate post-translational modification of SSG protein. Methods to detect such lesions in the SSGs of this invention are known to those of skill in the art.

Assay Techniques

Assay techniques that can be used to determine levels of gene expression (including protein levels), such as SSG of the present invention, in a sample derived from a patient are well known to those of skill in the art. Such assay methods include, without limitation, radioimmunoassays, reverse transcriptase PCR (RT-PCR) assays, immunohistochemistry assays, *in situ* hybridization assays, competitive-binding assays, Western Blot analyses, ELISA assays and proteomic approaches: two-dimensional gel electrophoresis (2D electrophoresis) and non-gel based approaches such as mass spectrometry or protein interaction profiling. Among these, ELISAs are frequently preferred to diagnose a gene's expressed protein in biological fluids.

An ELISA assay initially comprises preparing an antibody, if not readily available from a commercial source, specific to SSG, preferably a monoclonal antibody. In addition a reporter antibody generally is prepared which binds specifically to SSG. The reporter antibody is attached to a detectable reagent such as radioactive, fluorescent or enzymatic reagent, for example horseradish peroxidase enzyme or alkaline phosphatase.

To carry out the ELISA, antibody specific to SSG is incubated on a solid support, e.g. a polystyrene dish, that binds the antibody. Any free protein binding sites on the dish are then covered by incubating with a non-specific protein such as bovine serum albumin. Next, the sample to be analyzed is incubated in the dish, during which time SSG binds to the specific antibody attached to the polystyrene dish. Unbound sample is washed out with buffer. A reporter antibody specifically directed to SSG and linked to a detectable reagent such as horseradish peroxidase is placed in the dish resulting in binding of the reporter antibody to any monoclonal antibody bound to SSG. Unattached reporter antibody is then washed out. Reagents for peroxidase activity, including a colorimetric substrate are then added to the dish. Immobilized peroxidase, linked to SSG antibodies, produces a colored reaction product. The amount of color developed in a given time period is proportional to the amount of SSG protein present in the sample. Quantitative results typically are obtained by reference to a standard curve.

A competition assay can also be employed wherein antibodies specific to SSG are attached to a solid support and labeled SSG and a sample derived from the host are passed over the solid support. The amount of label detected which is attached to the solid support can be correlated to a quantity of SSG in the sample.

Using all or a portion of a nucleic acid sequence of a SSG of the present invention as a hybridization probe, nucleic acid methods can also be used to detect SSG mRNA as a marker for stomach cancer. Polymerase chain reaction (PCR) and other nucleic acid methods, such as ligase chain reaction (LCR) and nucleic acid sequence based amplification (NASBA), can be used to detect malignant cells for diagnosis and monitoring of various malignancies. For example, reverse-transcriptase PCR (RT-PCR) is a powerful technique which can be used to detect the presence of a specific mRNA population in a complex mixture of thousands of other mRNA species. In RT-PCR, an mRNA species is first reverse transcribed to complementary DNA (cDNA) with use of the enzyme reverse transcriptase; the cDNA is then amplified as in a standard PCR reaction. RT-PCR can thus reveal by amplification the presence of a single species of mRNA. Accordingly, if the mRNA is highly specific for the cell that produces it, RT-PCR can be used to identify the presence of a specific type of cell.

Hybridization to clones or oligonucleotides arrayed on a solid support (i.e. gridding) can be used to both detect the expression of and quantitate the level of expression of that gene. In this approach, a cDNA encoding the SSG gene is fixed to a substrate. The substrate may be of any suitable type including but not limited to glass, nitrocellulose, nylon or plastic. At least a portion of the DNA encoding the SSG gene is attached to the substrate and then incubated with the analyte, which may be RNA or a complementary DNA (cDNA) copy of the RNA, isolated from the tissue of interest. Hybridization between the substrate bound DNA and the analyte can be detected and quantitated by several means including but not limited to radioactive labeling or fluorescence labeling of the analyte or a secondary molecule designed to detect the hybrid. Quantitation of the level of gene expression can be done by comparison of the intensity of the signal from the analyte compared with that determined from known standards.

The standards can be obtained by *in vitro* transcription of the target gene, quantitating the yield, and then using that material to generate a standard curve.

Of the proteomic approaches, 2D electrophoresis is a technique well known to those in the art. Isolation of individual proteins from a sample such as serum is accomplished using sequential separation of proteins by different characteristics usually on polyacrylamide gels. First, proteins are separated by size using an electric current. The current acts uniformly on all proteins, so smaller proteins move farther on the gel than larger proteins. The second dimension applies a current perpendicular to the first and separates proteins not on the basis of size but on the specific electric charge carried by each protein. Since no two proteins with different sequences are identical on the basis of both size and charge, the result of a 2D separation is a square gel in which each protein occupies a unique spot. Analysis of the spots with chemical or antibody probes, or subsequent protein microsequencing can reveal the relative abundance of a given protein and the identity of the proteins in the sample.

The above tests can be carried out on samples derived from a variety of cells, bodily fluids and/or tissue extracts such as homogenates or solubilized tissue obtained from a patient. Tissue extracts are obtained routinely from tissue biopsy and autopsy material. Bodily fluids useful in the present invention include blood, urine, saliva or any other bodily secretion or derivative thereof. By blood it is meant to include whole blood, plasma, serum or any derivative of blood.

In Vivo Targeting of SSG/Stomach Cancer Therapy

Identification of these SSGs is also useful in the rational design of new therapeutics for imaging and treating cancers, and in particular stomach cancer. For example, in one embodiment, antibodies which specifically bind to a SSG

can be raised and used *in vivo* in patients suspected of suffering from stomach cancer. Antibodies which specifically bind SSG can be injected into a patient suspected of having stomach cancer for diagnostic and/or therapeutic purposes.

5 Thus, another aspect of the present invention provides for a method for preventing the onset and treatment of stomach cancer in a human patient in need of such treatment by administering to the patient an effective amount of antibody. By "effective amount" it is meant the amount or concentration
10 of antibody needed to bind to the target antigens expressed on the tumor to cause tumor shrinkage for surgical removal, or disappearance of the tumor. The binding of the antibody to the overexpressed SSG is believed to cause the death of the cancer cell expressing such SSG. The preparation and use of
15 antibodies for *in vivo* diagnosis and treatment is well known in the art. For example, antibody-chelators labeled with Indium-111 have been described for use in the radioimmunoscentigraphic imaging of carcinoembryonic antigen expressing tumors (Sumerdon et al. Nucl. Med. Biol. 1990
20 17:247-254). In particular, these antibody-chelators have been used in detecting tumors in patients suspected of having recurrent colorectal cancer (Griffin et al. J. Clin. Onc. 1991 9:631-640). Antibodies with paramagnetic ions as labels for use in magnetic resonance imaging have also been described
25 (Lauffer, R.B. Magnetic Resonance in Medicine 1991 22:339-342). Antibodies directed against SSG can be used in a similar manner. Labeled antibodies which specifically bind SSG can be injected into patients suspected of having stomach cancer for the purpose of diagnosing or staging of the disease
30 status of the patient. The label used will be selected in accordance with the imaging modality to be used. For example, radioactive labels such as Indium-111, Technetium-99m or Iodine-131 can be used for planar scans or single photon emission computed tomography (SPECT). Positron emitting
35 labels such as Fluorine-19 can be used in positron emission

tomography. Paramagnetic ions such as Gadlinium (III) or Manganese (II) can be used in magnetic resonance imaging (MRI). Presence of the label, as compared to imaging of normal tissue, permits determination of the spread of the cancer. The amount of label within an organ or tissue also allows determination of the presence or absence of cancer in that organ or tissue.

Antibodies which can be used in *in vivo* methods include polyclonal, monoclonal and omniclonal antibodies and antibodies prepared via molecular biology techniques. Antibody fragments and aptamers and single-stranded oligonucleotides such as those derived from an *in vitro* evolution protocol referred to as SELEX and well known to those skilled in the art can also be used.

15 **Vaccines**

Another aspect of the invention relates to compositions and methods for inducing an immunological response in a mammal. In one embodiment, a mammal is inoculated with a SSG polypeptide, or a fragment or variant thereof, in an amount adequate to produce an antibody and/or T cell immune response against SSG polypeptide. In another embodiment, a vector directing expression of SSG polynucleotide *in vivo* is used to induce such an immunological response and to produce antibody. The immune response against the SSG polypeptide is expected to protect the mammal from diseases, in particular stomach cancer.

Thus, the present invention also relates to an immunological/vaccine formulation (composition) which, when introduced into a mammal, induces an immunological response in that mammal to SSG polypeptide wherein the composition comprises a SSG polypeptide, fragment or variant thereof or a vector expressing a SSG gene or fragment thereof. The vaccine formulation may further comprise a suitable carrier. Since SSG polypeptide may be broken down in the stomach, the vaccine formulation is preferably administered parenterally

via subcutaneous, intramuscular, intravenous, or intradermal injection. Formulations suitable for parenteral administration include aqueous and non-aqueous sterile injection solutions which may contain anti-oxidants, buffers, bacteriostats and solutes which render the formulation isotonic with the blood of the recipient; and aqueous and non-aqueous sterile suspensions which may include suspending agents or thickening agents. The formulations may be presented in unit-dose or multi-dose containers, for example, as sealed ampules and vials, and may be stored in a freeze-dried condition requiring only the addition of the sterile liquid carrier immediately prior to use. The vaccine formulation may also include adjuvant systems for enhancing the immunogenicity of the formulation, such as oil-in water systems and other systems known in the art. The dosage will depend on the specific activity of the vaccine and can be readily determined by routine experimentation.

Screening Assays

The SSG polypeptides of the present invention can also be employed in screening processes for compounds which activate (agonists) or inhibit activation of (antagonists, or otherwise called inhibitors) the SSG polypeptides of the present invention. Thus, polypeptides of the invention can be used to identify agonists or antagonists from, for example, cells, cell-free preparations, chemical libraries, and natural product mixtures. These agonists or antagonists may be natural or modified substrates, ligands, receptors, enzymes, etc., as the case may be, of the polypeptides of the present invention; or may be structural or functional mimetics of the polypeptide of the present invention. See Coligan et al., *Current Protocols in Immunology* 1(2):Chapter 5 (1991).

SSG polypeptides are responsible for various biological functions, including pathologies such as stomach cancer. Accordingly, it is desirable to identify compounds which stimulate SSG polypeptides on the one hand (agonists) and

which can inhibit the function of SSG polypeptides (antagonists) on the other hand. Agonists and antagonists can be employed for therapeutic and prophylactic purposes for conditions such as stomach cancer.

5 In general, such screening procedures involve using appropriate cells which express the SSG polypeptide or respond to SSG polypeptide of the present invention. Such cells include those from mammals, yeast, *Drosophila* and *E. coli*. Cells which express the SSG polypeptide (or cell membranes
10 containing the expressed polypeptide) or respond to SSG polypeptides are then contacted with a candidate compound to observe binding, or stimulation or inhibition of a functional response. The SSG activity of the cells which were contacted with the candidate compounds is compared with the SSG activity
15 in the same type of cells which were not contacted with the candidate compounds.

The assays may simply test binding of a candidate compound wherein adherence to the cells bearing the SSG polypeptide is detected by means of a label directly or
20 indirectly associated with the candidate compound or in an assay involving competition with a labeled competitor. Further, these assays can test whether the candidate compound results in a signal generated by activation of the SSG polypeptide using detection systems appropriate to the cells
25 bearing the SSG polypeptide. Inhibitors of activation are generally assayed in the presence of a known agonist and the effect of the candidate compound upon activation by the agonist is observed.

Alternatively, the assays may comprise the steps of
30 mixing a candidate compound with a solution containing a SSG polypeptide to form a mixture, measuring SSG activity in the mixture, and comparing the SSG activity of the mixture to a standard.

The SSG polynucleotide, polypeptides and antibodies
35 immunospecific for the polypeptides can also be used to

configure assays for detecting the effect of added compounds on the production of SSG mRNA and polypeptides in cells. For example, an ELISA for measuring secreted or cell associated levels of SSG polypeptide using monoclonal and polyclonal
5 antibodies can be constructed by standard methods known in the art. The ELISA can then be used to discover agents which may inhibit or enhance the production of SSG from suitably manipulated cells or tissues. Standard methods for conducting these screening assays are well understood in the art.

10 The SSG polypeptides can also be used to identify membrane bound or soluble receptors, if any, through standard receptor binding techniques known in the art. These include, but are not limited to, ligand binding and crosslinking assays in which the SSG is labeled with a radioactive isotope (e.g.
15 ¹²⁵I), chemically modified (e.g. biotinylated), or fused to a peptide sequence suitable for detection or purification, and incubated with a source of the putative receptor (cells, cell membranes, cell supernatants, tissue extracts, bodily fluids). Other methods include biophysical techniques such as surface
20 plasmon resonance and spectroscopy. In addition to being used for purification and cloning of the receptor, these binding assays can be used to identify agonists and antagonists of SSG which compete with the binding of SSG to receptors. Standard methods for conducting these screening assays are well
25 understood in the art.

Examples of potential SSG polypeptide antagonists include, but are not limited to: antibodies; oligonucleotides or proteins which are closely related to the SSGs; ligands, substrates, receptors, and enzymes of the SSG polypeptides;
30 fragment of these ligands, substrates, receptors and enzymes; and small molecules which bind to the polypeptide of the present invention so that the activity of the polypeptide is prevented.

Thus, the present invention also relates to screening
35 kits for identifying agonists, antagonists, ligands,

receptors, substrates, enzymes, etc. for SSG polypeptides; or compounds which decrease or enhance the production of SSG polypeptides. Such kits preferably comprise a SSG polypeptide; a recombinant cell expressing a SSG polypeptide
5 or a cell membrane expressing a SSG polypeptide; and an antibody to a SSG polypeptide.

Prophylactic and Therapeutic Methods

This invention also relates to methods of treating abnormal conditions such as, stomach cancer, related to both
10 an excess of and insufficient amounts of SSG polypeptide activity.

If the activity of SSG polypeptide is in excess, several approaches are available. One approach comprises administering to a subject an inhibitor compound (antagonist)
15 as hereinabove described along with a pharmaceutically acceptable carrier in an amount effective to inhibit the function of the SSG polypeptide, such as, for example, by blocking the binding of ligands, substrates, enzymes, receptors, etc., or by inhibiting a second signal, and
20 thereby alleviating the abnormal condition. In another approach, soluble forms of SSG polypeptides still capable of binding the ligand, substrate, enzymes, receptors, etc. in competition with endogenous SSG polypeptide can be administered. Typical embodiments of such competitors
25 comprise fragments of the SSG polypeptide.

In still another approach, expression of the gene encoding endogenous SSG polypeptide can be inhibited using expression blocking techniques. Known blocking techniques involve the use of antisense sequences, either internally
30 generated or separately administered. See, for example, O'Connor, *J Neurochem* (1991) 56:560 in Oligodeoxynucleotides as Antisense Inhibitors of Gene Expression, CRC Press, Boca Raton, FL (1988). Alternatively, oligonucleotides which form triple helices with the gene can be supplied. See, for
35 example, Lee et al., *Nucleic Acids Res* (1979) 6:3073; Cooney

et al., *Science* (1988) 241:456; Dervan et al., *Science* (1991) 251:1360. These oligomers can be administered *per se* or the relevant oligomers can be expressed *in vivo*.

Several approaches are also available for treating
5 abnormal conditions related to an under-expression of SSG and its activity. One approach comprises administering to a subject a therapeutically effective amount of a compound which activates SSG polypeptide, i.e., an agonist as described above, in combination with a pharmaceutically acceptable
10 carrier, to thereby alleviate the abnormal condition. Alternatively, gene therapy can be employed to effect the endogenous production of SSG by the relevant cells in the subject. For example, a polynucleotide of the invention can be engineered for expression in a viral vector such as a
15 replication defective retroviral vector. The retroviral expression construct can then be isolated and introduced into a packaging cell transduced with a retroviral plasmid vector containing RNA encoding a polypeptide of the present invention such that the packaging cell now produces infectious viral
20 particles containing the gene of interest. These producer cells can be administered to a subject for engineering cells *in vivo* and expression of the polypeptide *in vivo*. For an overview of gene therapy, see Chapter 20, *Gene Therapy and other Molecular Genetic-based Therapeutic Approaches*, (and
25 references cited therein) in *Human Molecular Genetics*, T Strachan and A P Read, BIOS Scientific Publishers Ltd (1996). Another approach is to administer a therapeutic amount of SSG polypeptides in combination with a suitable pharmaceutical carrier.

30 **Formulation and Administration**

Peptides, such as a soluble form of SSG polypeptide, and agonist and antagonist peptides or small molecules, can be formulated in various combinations with suitable pharmaceutical carriers. These formulations comprise a

therapeutically effective amount of the peptide or small molecule, and a pharmaceutically acceptable carrier or excipient. Such carriers include, but are not limited to, saline, buffered saline, dextrose, water, glycerol, ethanol, and combinations thereof. The formulation is selected in accordance with the mode of administration, and is well within the skill of the art. The invention further relates to pharmaceutical packs and kits comprising one or more containers filled with one or more of the ingredients of the
10 aforementioned compositions of the invention.

The compositions of present invention can be employed alone or in conjunction with other compounds, such as other therapeutic compounds.

Preferred forms of systemic administration of these
15 pharmaceutical compositions include injection, typically by intravenous injection. Other injection routes, such as subcutaneous, intramuscular, or intraperitoneal, can also be used. Alternative means for systemic administration include transmucosal and transdermal administration using penetrants
20 such as bile salts or fusidic acids or other detergents. In addition, if properly formulated in enteric or encapsulated formulations, oral administration may also be possible. These compositions can also be administered topically in the form of salves, pastes, gels and the like.

25 The dosage range required depends on the composition, the route of administration, the nature of the formulation, the nature of the subject's condition, and the judgment of the attending practitioner. Suitable dosages, however, are generally in the range of 0.1-100 μ g of peptide or small
30 molecule per kg of subject. Wide variations in the needed dosage, however, are to be expected in view of the variety of compounds available and the differing efficiencies of various routes of administration. For example, oral administration is expected to require higher dosages than administration by
35 intravenous injection. Variations in these dosage levels can

in which the differentially expressed messages are sought and Driver: cDNA population in which the differentially expressed transcripts are absent or low). The tester sample is separated in two parts and different PCR adapters are ligated to the 5' ends. Each tester is separately annealed to excess driver (first annealing) and then pooled and again annealed (second annealing) to excess driver. During the first annealing, sequences common to both populations anneal. Additionally, the concentration of high and low abundance messages are normalized since annealing is faster for abundant molecules due to the second order kinetics of hybridization. During the second annealing, cDNAs unique or overabundant to the tester can anneal together. Such molecules have different adapters at their ends. The addition of additional driver during the second annealing enhances the enrichment of the desired differentially expressed sequences. During subsequent PCR, molecules that have different adapters at each end amplify exponentially. Molecules which have identical adapters, or adapters at only one end, or no adapters (driver sequences) either do not amplify or undergo linear amplification. The end result is enrichment for cDNAs corresponding to differentially expressed messages (unique to the tester or upregulated in the tester).

This technique was used to identify transcripts unique to stomach tissues or messages overexpressed in stomach cancer. Pairs of matched samples isolated from the same patient, a cancer sample, and the "normal" adjacent tissue from the same tissue type were utilized. The mRNA from the cancer tissue is used as the "tester", and the non-cancer mRNA as a "driver". The non-cancer "driver" is from the same individual and tissue as the cancer sample (Matched). Alternatively the "driver" can be from a different individual but the same tissue as the tumor sample (unmatched). In some cases, mixtures of mRNAs derived from non-cancer tissue types different from the cancer tissue type were used as the

"driver". This approach allows the identification of transcripts whose expression is specific or upregulated in the cancer tissue type analyzed.

Several subtracted libraries were generated for stomach.

5 The product of the subtraction experiments was used to generate cDNA libraries. These cDNA libraries contain Expressed Sequence Tags (ESTs) from genes that are stomach cancer specific, or upregulated in stomach. Selected clones from each cDNA PCR Select library were sequenced and are

10 depicted as SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 or 13.

Example 2: Relative Quantitation of Gene Expression

Real-Time quantitative PCR with fluorescent Taqman probes is a quantitation detection system utilizing the 5'-3' nuclease activity of Taq DNA polymerase. The method uses an

15 internal fluorescent oligonucleotide probe (Taqman) labeled with a 5' reporter dye and a downstream, 3' quencher dye. During PCR, the 5'-3' nuclease activity of Taq DNA polymerase releases the reporter, whose fluorescence can then be detected

20 by the laser detector of the Model 7700 Sequence Detection System (PE Applied Biosystems, Foster City, CA, USA).

Amplification of an endogenous control is used to standardize the amount of sample RNA added to the reaction and normalize for Reverse Transcriptase (RT) efficiency. Either

25 use cyclophilin, glyceraldehyde-3-phosphate dehydrogenase (GAPDH) or 18S ribosomal RNA (rRNA) were used as this endogenous control. To calculate relative quantitation between all the samples studied, the target RNA levels were used for one sample as the basis for comparative results

30 (calibrator). Quantitation relative to the "calibrator" can be obtained using the standard curve method or the comparative method (User Bulletin #2: ABI PRISM 7700 Sequence Detection System).

The tissue distribution and the level of the target gene for every example in normal and cancer tissue were evaluated. Total RNA was extracted from normal tissues, cancer tissues, and from cancers and the corresponding matched adjacent
5 tissues. Subsequently, first strand cDNA was prepared with reverse transcriptase and the polymerase chain reaction was done using primers and Taqman probe specific to each target gene. The results are analyzed using the ABI PRISM 7700 Sequence Detector. The absolute numbers are relative levels
10 of expression of the target gene in a particular tissue compared to the calibrator tissue.

Leads evaluated using real-time quantitative PCR are named "sto" followed by a number (for example: sto108).

Example 3: Comparative Examples

15 Similar mRNA expression analysis for genes coding for the diagnostic markers PSA (Prostate Specific Antigen) and PLA2 (Phospholipase A2) were performed for comparison. PSA is the only cancer screening marker available in clinical laboratories. When the panel of normal pooled tissues was
20 analyzed, PSA was expressed at very high levels in prostate, with a very low expression in breast and testis. After analysis of more than 55 matching samples from 14 different tissues, the data corroborated the tissue specificity seen with normal tissue samples. PSA expression was compared in
25 cancer and normal adjacent tissue for 12 matching samples of prostate tissue. The relative levels of PSA were higher in 10 cancer samples (83%). Clinical data recently obtained support the utilization of PLA2 as a staging marker for late stages of prostate cancer. This mRNA expression data showed
30 overexpression of the mRNA in 8 out of the 12 prostate matching samples analyzed (66%). The tissue specificity for PLA2 was not as good as the one described for PSA. In addition to prostate, small intestine, liver, and pancreas also showed high levels of mRNA expression for PLA2.

**Example 4: Semi-quantitative Polymerase Chain Reaction
 (SQ-PCR)**

SQ-PCR is a method that utilizes end point PCR on serial dilutions of cDNA samples in order to determine relative expression patterns of genes of interest in multiple samples. Using random hexamer primed Reverse Transcription (RT) cDNA panels are created from total RNA samples. Gene specific primers are then used to amplify fragments using Polymerase Chain Reaction (PCR) technology from four 10x serial cDNA dilutions in duplicate. Relative expression levels of 0, 1, 10, 100 and 1000 are used to evaluate gene expression. A positive reaction in the most dilute sample indicates the highest relative expression value. This is determined by analysis of the sample reactions on a 2-4% agarose gel. The tissue samples used include 12 normal, 12 cancer and 6 pairs of tissue specific cancer and matching samples. Leads evaluated through this method are named sqsto followed by a number (for example: sqsto002)

SEQ ID NO:1; Clone ID sto342571F1 (Sto108/sqsto002)

Real-Time quantitative PCR was done using the following primers:

Sto108 forward:

AGGGGAAGGGCTAGAACAAAAT (SEQ ID NO:14)

Sto108 reverse:

AGCATGAGTGGACCTACACAAGA (SEQ ID NO:15)

Q-PCR probe

TAAGGTCATGTGGTCTCTGCCTGCTCTCT (SEQ ID NO:16)

Table 1 shows the absolute numbers which are relative levels of expression of Sto108 in 24 normal samples from different tissues. All the values are compared to normal testis (calibrator). These RNA samples are commercially

available pools, originated by pooling samples of a particular tissue from different individuals.

Table 1: Stot108 in Pooled Tissue Samples

	Tissue	Normal
5	Adrenal Gland	0.01
	Bladder	0.00
	Brain	0.02
	Cervix	0.00
	Colon	0.00
10	Endometrium	0.00
	Esophagus	0.00
	Heart	0.00
	Kidney	0.00
	Liver	0.06
15	Lung	0.00
	Mammary gland	0.00
	Muscle	0.01
	Ovary	0.00
	Pancreas	0.00
20	Prostate	0.10
	Rectum	0.00
	Small Intestine	0.00
	Spleen	0.23
	Stomach	0.08
25	Testis	1.00
	Thymus	0.00
	Trachea	0.00
	Uterus	0.00

0.00= Negative

30 The relative levels of expression in Table 1 show that Stot108 mRNA expression is detected in the pool of a few normal tissue analyzed.

The absolute numbers in Table 1 were obtained analyzing pools of samples of a particular tissue from different
35 individuals. They can not be compared to the absolute numbers originated from RNA obtained from tissue samples of a single individual in Table 2.

Table 2 shows the absolute numbers which are relative levels of expression of Stot108 in 20 pairs of matching
40 samples. All the values are compared to normal testis (calibrator). A matching pair is formed by mRNA from the cancer sample for a particular tissue and mRNA from the normal adjacent sample for that same tissue from the same individual.

Table 2: Sto108 in Individual Tissue Samples

	Sample ID	Tissue	Cancer	Normal Adjacent Tissue	Stage
	Sto542S	Stomach 1	0.00	0.00	IB
	StoAC99	Stomach 2	0.31	0.41	IB
5	Sto915S	Stomach 3	0.00	0.00	II
	Sto264S	Stomach 4	0.54	0.00	II
	Sto758S	Stomach 5	0.16	0.00	IIIA
	Sto17S	Stomach 6	0.00	2.09	IIIA
	Sto27S	Stomach 7	0.00	0.76	IIIB
10	Sto539S	Stomach 8	0.00	0.00	IIIB
	Sto490S	Stomach 9	0.00	0.00	IV
	Sto728	Stomach 10	0.00	0.00	NA
	ClnDC63	Colon	0.26	0.25	
	LngSQ80	Lung	0.20	0.26	
15	Endo12XA	Endometrium	0.16	0.00	
	Mam986	Mammary gland	0.21	0.00	
	Kid5XD	Kidney	0.00	0.00	
	Pan71XL	Pancreas	0.00	0.52	
	SmIntH89	Small Intestine	0.00	0.00	
20	Thr270T	Thyroid 1	0.00	0.00	
	Thr590D	Thyroid 2	0.00	0.17	
	Pro326	Prostate	0.22	0.00	

0.00= Negative

NA= information is not available

25 Table 2 represents 40 samples in 10 different tissues. Table 1 and Table 2 represent a combined total of 64 samples in 25 human tissue types. Comparisons of the level of mRNA expression in cancer samples and the normal adjacent tissue from the same individuals are shown in Table 2. Sto108 is

30 expressed at higher levels in 2 of 10 (20%) cancer samples (stomach 4 & 5) compared to normal adjacent tissue.

SEQ ID NO:2; Clone ID sto347444F1 (Sto109/sqsto010)

Real-Time quantitative PCR was done using the following primers:

35 Sto109 forward:

ATGGCTTACAGTCCCGAGGA (SEQ ID NO:17)

Sto109 reverse:

CTCCCTGATCTGTACACCAAC (SEQ ID NO:18)

Q-PCR probe:

TCGCCCCACTTCTACTGCCTTGTCTTC (SEQ ID NO:19)

Table 3 shows the absolute numbers which are relative levels of expression of Stol09 in 24 normal samples from different tissues. All the values are compared to normal muscle (calibrator). These RNA samples are commercially available pools, originated by pooling samples of a particular tissue from different individuals.

Table 3: Stol09 in Pooled Tissue Samples

	Tissue	Normal
10	Adrenal Gland	3.26
	Bladder	0.00
	Brain	8.34
	Cervix	5.68
	Colon	0.00
15	Endometrium	10.93
	Esophagus	0.97
	Heart	0.60
	Kidney	2.44
	Liver	11.24
20	Lung	44.79
	Mammary gland	1.97
	Muscle	1.00
	Ovary	18.44
	Pancreas	14.98
25	Prostate	8.60
	Rectum	8.91
	Small Intestine	2.74
	Spleen	41.93
	Stomach	3.85
30	Testis	14.12
	Thymus	49.69
	Trachea	8.31
	Uterus	11.55

0.00= Negative

35 The relative levels of expression in Table 3 show that Stol09 mRNA expression is detected in the pool of a few normal tissue analyzed.

The absolute numbers in Table 3 were obtained analyzing pools of samples of a particular tissue from different individuals. They can not be compared to the absolute numbers originated from RNA obtained from tissue samples of a single individual in Table 4.

Table 4 shows the absolute numbers which are relative levels of expression of Sto109 in 40 pairs of matching samples. All the values are compared to normal muscle (calibrator). A matching pair is formed by mRNA from the 5 cancer sample for a particular tissue and mRNA from the normal adjacent sample for that same tissue from the same individual.

Table 4: Sto109 in Individual Tissue Samples

	Sample ID	Tissue	Cancer	Normal Adjacent Tissue	Stage
	Sto542S	Stomach 1	25.02	29.34	I
10	Sto264S	Stomach 2	21.11	11.84	II
	Sto915S	Stomach 3	6.08	1.63	II
	Sto115S	Stomach 4	20.32	5.05	III
	Sto15S	Stomach 5	5.74	9.92	III
	Sto261S	Stomach 6	7.29	3.69	III
15	Sto17S	Stomach 7	24.17	8.91	III
	Sto27S	Stomach 8	9.32	23.10	III
	StoAC99	Stomach 9	16.39	14.52	I
	Sto758S	Stomach 10	9.45	3.85	III
	Sto539S	Stomach 11	3.28	10.34	III
20	Sto531S	Stomach 12	15.08	4.00	IV
	Sto490S	Stomach 13	9.68	14.03	IV
	Sto288S	Stomach 14	12.13	4.82	NA
	Sto728	Stomach 15	32.11	2.63	NA
	Bld46XK	Bladder 1	2.08	1.41	
25	BldTR14	Bladder 2	20.89	36.50	
	ClnCM67	Colon 1	4.21	3.68	
	ClnDC19	Colon 2	28.05	13.32	
	ClnDC63	Colon 3	22.09	12.68	
	Endo10479	Endometrium 1	38.99	21.56	
30	Endo12XA	Endometrium 2	28.05	19.56	
	Kid5XD	Kidney 1	3.05	16.97	
	Liv15XA	Liver 1	6.43	3.45	
	LngSQ43	Lung 1	21.93	4.45	
	LngSQ9X	Lung 2	35.51	11.24	
35	Lng143L	Lung 3	13.32	3.42	
	Lng77L	Lung 4	2.25	10.56	
	LngAC39	Lung 5	11.67	79.62	
	LngSQ80	Lung 6	40.22	18.83	
	Mam986	Mammary 1	5.56	6.61	
40	Pan82XP	Pancreas 1	5.98	0.00	
	Pan 71XL	Pancreas 2	9.00	9.38	
	Pro326	Prostate 1	8.75	3.53	
	SmInt21XA	Small Intestine 1	6.17	0.59	
	SmIntH89	Small Intestine 2	6.19	7.04	
45	Thr590D	Thyroid 1	27.00	9.32	
	Thr270T	Thyroid 2	34.30	51.63	

Tst647T	Testis 1	11.59	23.26	
Utr135XO	Uterus 1	19.23	20.11	

0.00= Negative

NA= information is not available

5 Table 4 represents 80 samples in 14 different tissues.
Table 3 and Table 4 represent a combined total of 104 samples
in 25 human tissue types. Comparisons of the level of mRNA
expression in cancer samples and the normal adjacent tissue
from the same individuals are shown in Table 4. Sto109 is
10 expressed at higher levels in 10 of 15 (67%) cancer samples
(stomach 2-4, 6-7, 9-10,12, 14-15) compared to normal adjacent
tissue.

TABLE 4: TESTES